

## **CORE Leptospirosis**

Rapid test for the detection of IgM antibodies to Leptospira  
(Device)

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### **INTRODUCTION**

Core Leptospirosis is a rapid, self-performing, qualitative, sandwich immunoassay for the detection of Leptospira specific IgM antibodies in human serum/plasma or whole blood specimen. It is useful for the serodiagnosis of current or recent Leptospirosis. The broadly reactive genus specific antigen employed in the test allows the detection of Leptospira infections caused by a wide range of strains of different serovars.

### **SUMMARY**

Leptospira are actively motile, delicate spirochaetes possessing a large number of closely wound spirals and characteristic hooked ends. There are several species of Leptospira and they may be saprophytic or parasitic. They can be distinguished only under dark ground illumination in the living state or by electron microscopy. Leptospirosis is a zoonotic disease of worldwide prevalence. Humans are infected when the water contaminated by the urine of carrier animals enters the body through cuts or abrasions on the skin or through intact mucosa of the mouth, nose or conjunctiva. Clinical symptoms include fever, chills, headache, conjunctivitis, myalgia and GI related symptoms, Kidney infection is a common sequelae.

Diagnosis may be made by demonstration of Leptospire microscopically in blood or urine, by isolating them in culture or by inoculation of guinea pigs, or by serological tests.

Core Leptospirosis, qualitatively detects the presence of IgM class of Leptospira specific antibodies in human serum /plasma or whole blood specimen.

### **PRINCIPLE**

Core Leptospirosis-WB utilizes the principle of immunochromatography, a unique two-site immunoassay on a membrane. As the test sample flows through the membrane assembly of the test device, the anti human IgM -colloidal gold conjugate forms a complex with IgM antibodies in the sample. This complex moves further on the membrane to the test window 'T' where it is immobilized by the broadly reactive Leptospira genus specific antigens coated on the membrane, leading to the formation of a red to deep purple coloured band at the test region 'T' which confirms a positive test result. Absence of this coloured band in test region 'T' indicates a negative test result. The unreacted conjugate and the unbound complex if any move further on the membrane and are subsequently immobilized by the anti-rabbit antibodies coated on the control window 'C' of the membrane assembly, forming a red to deep purple coloured band. The control band serves to validate the test results.

### **REAGENTS AND MATERIALS SUPPLIED**

Each kit contains:

- A. Individual pouches, each containing:
  1. Test Device: Membrane test assembly predisposed with Anti Human IgM -colloidal gold conjugate, Leptospira genus specific antigens at test window 'T' and anti- rabbit antiserum predisposed at the control window 'C'.
  2. Desiccant pouch
  3. 5µl sample loops.
- B. Sample Running Buffer
- C. Package Insert

### **STORAGE AND STABILITY**

The sealed pouches in the test kit & the kit components may be stored between 4-30°C for the duration of the shelf life as indicated on the pouch.

**OPTIONAL MATERIAL REQUIRED:** 10µl precision micropipettes.

### **NOTE**

1. For in vitro diagnostic use only. NOT FOR MEDICINAL USE.
2. Do not use beyond expiry date.
3. Read the instructions carefully before performing the test.
4. Handle all specimens as potentially infectious.
5. Follow standard biosafety guidelines for handling and disposal of potentially infective material.

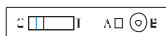
### **SPECIMEN COLLECTION AND PREPARATION**

1. Blood samples collected with a suitable anticoagulant such as EDTA or Heparin or Oxalate can also be used.
2. No prior preparation of the patient is required before sample collection by approved techniques.
3. Fresh serum / plasma is preferable. Anticoagulated whole blood can also be used as specimen. Serum / plasma may be stored at 2-8°C up to 24 hours in case of delay in testing. For long-term storage, freeze the specimen at -20°C for 3 months or -70°C for longer periods. Whole blood should be used immediately and should not be frozen.
4. Repeated freezing and thawing of the specimen should be avoided.
5. Do not use haemolysed, clotted, contaminated, viscous/turbid specimen.
6. Specimen containing precipitates or particulate matter must be centrifuged and the clear supernatant only used for testing.
7. For each sample a new sample loop should be used.

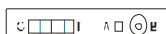
### **TESTING PROCEDURE AND INTERPRETATION OF RESULTS**

1. Bring the kit components to room temperature before testing.
2. Open the pouch and retrieve the test device. Once opened, the device must be used immediately.
3. Label the test device with the patient's identity.
4. Add 10µl of serum/ plasma or whole blood with a micropipette into the sample port "A", OR using the 5µl sample loop provided with the kit. Dip the loop into the sample and then blot into the sample port 'A'. Repeat this step twice for each sample. Ensure that the loop does not retrieve clots or debris from the sample.
5. Add 5 drops of sample running buffer to the reagent port "B"
6. At the end of 15 minutes read the results as follows.

Negative Test Result:



Positive Test Result:



7. The test should be considered invalid if neither the control band 'C' nor the test band 'T' appears. Repeat the test with a new device.

### **Performance Characteristics**

Core Leptospirosis - WB was evaluated at the Royal Tropical Institute, Amsterdam in parallel with other licensed tests for the serodiagnosis of Leptospirosis. The 47 sera evaluated were from diverse serogroups of *Leptospira*. Core Leptospirosis-WB had a performance comparable to the other tests.

#### Remarks:

1. The intensity of the test line depends upon the stage of the disease and the titres of the antibodies in the test specimen.
2. As specific antibodies reach detectable levels about one week after the onset of disease, a sample collected very early may yield a negative test result.
3. If the test is negative and if Leptospirosis is still suspected the test should be repeated with the second sample collected at a later date in conjunction with clinical reexamination.
4. In endemic areas faint bands may appear occasionally due to borderline IgM titres present as a result of previous exposures.
5. It is recommended that the positive results obtained must be reconfirmed using a confirmatory test such as the MAT (Microscopic agglutination test).
6. High titres of RF and heterophile antibodies may interfere with the test, in such cases the results must be interpreted with caution.
7. The results must be correlated with clinical findings to arrive at the diagnosis.
8. Do not use the test kit beyond expiration date.

#### Bibliography

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